

SELECTIVE INTRACOMPLEX REACTION OF PENDANT GROUPS IN A PURINE-PYRIMIDINES BASE PAIR

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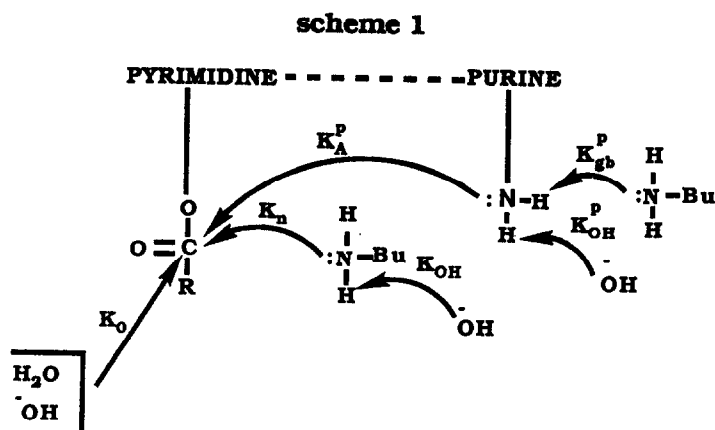
Abstract: Evidence is presented suggesting that hydrogen-bonded base pairing and/or aryl stacking interactions between derivatives of guanine (9-[(4-aminobutyloxy)-methyl]guanine) and cytosine (1-[(2-benzyloxy)ethoxy]methyl)cytosine) lead to enhanced intracomplex chemical reaction between the corresponding amino and ester groups. On the basis of analysis of the kinetic data it is concluded that a chain length of four methylene groups (4-amino-butyloxy as compared to 2-aminoethoxy or 6-aminohexyloxy) on the guanine is necessary to achieve the appropriate geometry for intracomplex reaction. Support for the reaction scheme is provided by the absence of reaction between the guanine derivative and a corresponding ester derivative of thymine, not expected to associate.

INTRODUCTION

One important feature affecting enzymatic reaction efficiency is believed to be the specificity of binding and precise orientation of the reacting species in the enzyme substrate complex. Based in part upon this view, the last two decades have seen the design of many synthetic models involving intramolecular reactions of covalently bound reactants⁽¹⁾ and bimolecular reactions resulting from specific binding interactions (electrostatic, hydrophobic) of reactants.^(2,3,4) Host and guest molecular recognition has also attracted much interest and selective hosts for anionic⁽⁵⁾ cationic⁽⁶⁾ and hydrophobic^(3a,7) guests have been synthesized.

The base pairing of purines and pyrimidines^(8a,8b) which is essential for the intramolecular attraction and recognition in nucleic acids would appear attractive as a model for bringing together two different potentially reactive groups for selective chemical reaction. Several approaches to the design of such model systems have recently been reported in the literature.^(8,9)

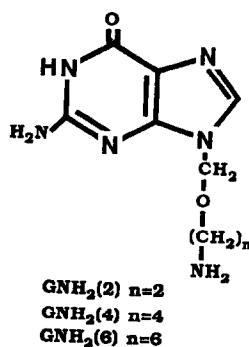
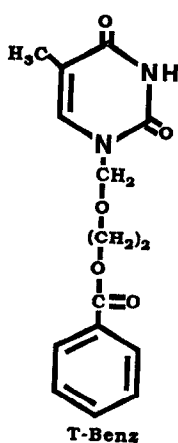
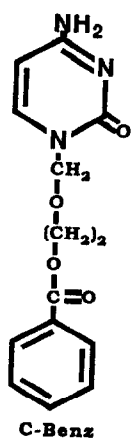
Here we report that hydrogen bonded base pairing and/or aryl stacking interactions between a guanine linked to an alkylamine and a cytosine linked to an alkylbenzoate lead to enhanced intra-complex reaction. The kinetic model in the presence of n-butylamine as buffer and an external catalyst is depicted in Scheme 1.



The dotted line denotes purine-pyrimidine coupled bases linked to an amine group and ester moiety, respectively. BuNH_2 and OH are external catalysts.

k_n and k_{OH} represent rate constants of nucleophilic attack by external amines and specific base catalyzed external amine attack on the ester site in the purine-pyrimidine pair, respectively. k_{gb}^{p} and k_{OH}^{p} however, denote the BuNH_2 and OH catalyzed nucleophilic attack of Purine- NH_2 in the ternary activated complex and k_{A}^{p} corresponds to the internal, uncatalyzed nucleophilic attack of Purine- NH_2 . k_0 includes the water and specific base rate constant.

The following esters and catalysts were prepared:



EXPERIMENTAL

The syntheses of C-Benz, T-Benz and GNH_2 ($n=2,4,6$) were performed by the chloromethyl alkylation according to described procedures^(10,11,12) and had physical properties in accord with the structures given.

1-[[2-(Benzoyloxy)ethoxy]methyl]cytosine, (C-Benz)

m.p.=185°C, 187-188°C lit⁹, ¹H-nmr: δ 3.7, 9.17 (m, 4H(CH₂CH₂)), δ 5.02 (s, 2H, (OCH₂N)), δ 5.88 (d, 1H (C5)), δ 7.02, 7.63 (m, 8H (NH₂, C6, A_x)).

M.S.: 289M⁺, 179, 141, 111, 105.

Anal. calc'd. for C₁₄H₁₅N₃O₄; C, 58.1; H, 5.25; N, 14.53; Found: C, 58.17; H, 5.41; N, 14.72.

1-[[2-(Benzoyloxy)ethoxy]methyl] thymine, (T-Benz)

m.p.=115°C, 115-116°C lit⁹, ¹H-nmr: δ 1.85, (s, 3H(CH₃)) δ 3.9, 4.7 (dt, 4H (CH₂CH₂)) δ 5.2 (s, 2H (OCH₂N)) δ 7.1-7.43 (m, 7H, (C(3), C(6), Ar)).

M.S.: 304M⁺, 227, 199, 184, 170, 155, 139, 125.

Anal. calc'd. for C₁₅H₁₆N₂O₅; C, 59.21; H, 5.3; N, 19.72; Found: C, 59.67; H, 5.25; N, 9.74.

Found: C, 59.67; H, 5.25; N, 9.74.

All the 9-[aminoalkoxy]methyl derivatives of guanine were prepared as their phthaloyl analogs. The phthaloyl protective groups were cleaved with hydrazine followed by 1M HCl.

9-[(2-Aminoethoxy) methyl]guanine.HCl (GNH₂(2)).

m.p. = 228-230°C; ¹H-nmr (d⁶-DMSO); δ 7.84 (s, 1H (C8)), δ 6.95 (br.s, 2H (Purine NH₂)), δ 5.4 (s, 2H (NCH₂O)), δ 3.55 (t, 2H (OCH₂)) δ 2.84 (t, 2H (CH₂N)).

M.S.: 225M⁺, 224, 208, 194, 180, 164, 150.

Analysis calc'd for the phthalimido derivative C₁₆H₁₄N₆O₄, C, 54.23; H, 3.95; N, 23.72; Found: C, 54.82; H, 3.8; N, 23.4.

9-[(4-aminobutyloxy)methyl]guanine.HCl (GNH₂(4)).

m.p. = 246°C; ¹H-nmr (d⁶-DMSO): δ 7.85 (s, 1H (C8)), δ 6.94 (br.s, 2H (Purine NH₂)), δ 5.4 (s, 2H (NCH₂O)), δ 3.81, 3.6 (m, 6H (CH₂CH₂CH₂)) δ 2.87 (t, 2H (CH₂N)).

M.S.: 254M⁺, 253, 239, 225, 211, 180, 164, 150.

Anal. calc'd for the phthalimido derivative C₁₈H₁₆N₆O₄. C, 56.54; H, 4.71; N, 21.98; Found: C, 56.74; H, 4.65; N, 21.59.

9-[(6-aminohexyloxy)methyl]guanine.HCl (GNH₂(6)).

m.p. = 250°C; ¹H-nmr (d⁶-DMSO): δ 7.85 (s, 1H (C8)), δ 6.95 (br.s, 2H (Purine NH₂)), δ 5.4 (s, 2H (NCH₂O)), 3.95-3.6 (m, 12H), 1.6 (s, 2H (aliphatic NH₂)).

Anal. calc'd for the phthalimido derivative C₂₀H₂₂N₆O₄. C, 58.53; H, 5.36; N, 20.48; Found: C, 58.72; H, 5.21; N, 20.22.

The kinetic measurements of C-Benz hydrolysis were carried out spectrophotometrically by monitoring the change in absorbance V_s time at 234nm in the presence of external buffer (catalyst): butylamine (0-0.6M) or K₂CO₃ (0-0.1M) at pH 10.34, T=35.4°C.

The concentrations of C-Benz and of GNH₂ in solution were both 1.1x10⁻³M.

RESULTS AND DISCUSSION

The rate expression in the presence of butylamine as external catalyst and buffer is given in equation 1.

$$k_{obs} = k_o + k_n [BuNH_2] + k_{OH} [^-OH] [BuNH_2] + k_{gb}^p [BuNH_2] + k_{OH}^p [^-OH] + k_A^p.$$

Since the values of the rate constants k_o , k_n and k_{OH} of the coupled C-Benz-GNH₂ ester are assumed to be close to the corresponding rates resulting from the isolated C-benz ester, the difference between the slopes of the two systems and intercepts of the two systems determined by the linear plot of $k_{obs} V_m [BuNH_2]$, should equal k_{gb}^p and $k_{OH}^p [^-OH] + k_A^p$ respectively.

With K₂CO₃ as buffer the observed rate constant obey equation 2:

$$k_{obs} = k_{hyd} + k_{OH}^p [OH] + k_A^p + k_- [CO_3^{--}] \quad (2)$$

Here again the difference between the intercepts of the coupled and uncoupled esters determined by plots of $k_{obs} V_m [CO_3^{--}]$ equal to $k_{OH}^p [OH] + k_A^p$.

Fig 1 exhibits the dependence of C-Benz on BuNH₂ in the presence and absence of GNH₂ (n=4) at pH=10.3 T=35.4°C. The slopes and intercept values of C-Benz in the presence of BuNH₂ or K₂CO₃ with and without the complementary base GNH₂ (n=2,4,6) are assembled in Table 1. The data of T-Benz in aqueous solution and of C-Benz in dioxane solution is also included.

From Table 1 it is inferred that the spatial arrangement of the nucleophilic amino group attached to the complementary base, relative to the ester carbonyl group is crucial for catalysis. Only chain length n=4 on GNH₂ is appropriate for catalysis; no catalysis was observed with GNH₂ when n=2 or 6.

The calculated rate constants for n=4 are: $k_{gb}^p = 8.10^{-3} M^{-1} \text{ min}^{-1}$ and $k_{OH}^p [OH] + k_A^p = 3.1 \times 10^{-3} \text{ min}^{-1}$ (Table 1, entry 1,2 and entry 3,4). Additional evidence for our model involving a specific oriented disposition requirements are: a) addition of 5M urea abolished catalysis, b) substrates which are not expected to form base pairs (T-Benz + GNH₂ (n=4)) do not exhibit internal catalysis (Table 1 entries 7,8).

In aqueous solution it seems most likely that base association^(1,3) involves a stacking arrangement. In organic solvents however base pairing due to hydrogen bonding probably takes place. We have therefore also examined the catalytic effect by GNH₂ in dioxane as a solvent (Table 1, entry 9,10).

In non-aqueous solvent the kinetic terms contributing to catalysis according to equation 1 are: k_o , k_n , k_{gb}^p and k_A^p .

From Table 1 it is inferred that C-Benz + BuNH₂ in the absence of the complementary base (GNH₂) does not display any substantial catalysis ($k_n=0$) however in the presence of GNH₂ (n=4), k_{gb}^p and k_A^p contribute significantly and have values of $2.4 \times 10^{-3} M^{-1} \text{ min}^{-1}$ and $2.8 \times 10^{-4} \text{ min}^{-1}$ respectively.

In conclusion, our results indicate that reaction selectivity may be brought about by purine-pyrimidine base stacking and or base pairing. Further studies involving more efficient recognition sites and examining additional reactions, in modified purine-pyrimidine bases are now called for.

Table 1

Slopes and intercepts calculated by equations 1 and 2 for C-Benz and T-Benz with and without the complementary base GNH_2 ($n=2,4,6$) $\text{pH}=10.3$, $T=35.4^\circ\text{C}$.

No.	Ester	amine	Buffer	Slope $\text{M}^{-1} \text{min}^{-1}$	intercept min^{-1}
1	C-Benz	$\text{GNH}_2(4)$	BuNH_2	13.2×10^{-3}	5.2×10^{-3}
2	C-Benz	-	BuNH_2	5.2×10^{-3}	2.1×10^{-3}
3	C-Benz	$\text{GNH}_2(4)$	K_2CO_3	1.35×10^{-3}	5.1×10^{-3}
4	C-Benz	-	K_2CO_3	1.42×10^{-3}	2.1×10^{-3}
5	C-Benz	$\text{GNH}_2(2)$	BuNH_2	2.36×10^{-3}	1.75×10^{-3}
6	C-Benz	$\text{GNH}_2(6)$	BuNH_2	3.1×10^{-3}	2.0×10^{-3}
7	T-Benz	-	BuNH_2	1.7×10^{-3}	1.75×10^{-3}
8	T-Benz	$\text{GNH}_2(4)$	BuNH_2	2.0×10^{-3}	1.83×10^{-3}
9	C-Benz ^a	$\text{GNH}_2(4)$	BuNH_2	2.4×10^{-3}	0.4×10^{-3}
10	C-Benz ^a	-	BuNH_2	0.0	0.12×10^{-3}

^a in dioxane.

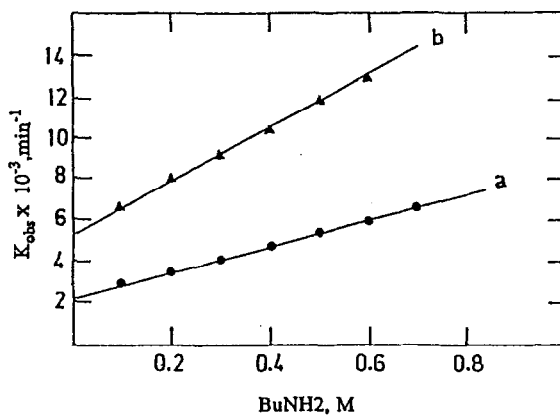


Fig. 1. First order rate constants of C-Benz V_s BuNH_2 concentration:
 a) Without adding $\text{GNH}_2(4)$ (-●-●-).
 b) With the adding of $\text{GNH}_2(4)$ (-▲-▲-), $\text{pH}=10.3$, $T=35.4^\circ\text{C}$.

REFERENCES

1. a) Bruice, T.C.; Benkovic, S.J. "Bioorganic Mechanisms" Vol. I, W.A. Benjamin, New York 1966, chapter 1-119. b) Jencks W.P. "Catalysis in Chemistry and Enzymology", McGraw Hill, New-York 1969, chapter 2, 42-162. c) M.C. Bender, R.J. Bergeron, M. Kamiyama "Bioorganic Chemistry of Enzymatic Catalysis" John Wiley, New York 1984, chapter 10.
2. a) Kunitake, T.; Shinkai, S. Adv. Phys. Org. Chem. 1980, 17, 435-487. b) J.M. Brown, in Colloid Science, Specialist periodical report, The Chemical Society, London, 1979 Vol. 3 253. c) Bunton, C.A. Catal. Rev. Sci. Eng. 1979 20 1. d) Fendler, J.H. , "Membrane Mimetic Chemistry" Wiley-Interscience, New York (1982).
3. a) Bender, M.L.; Komiyama, M. "Cyclodextrin Chemistry", Springer Verlag, New-York, 1978; b) Breslow, R. Science 1982, 218, 532. c) Tabushi, I. Acc. Chem. Res. 1982, 15, 66. d) D'Souza, V.T.; Bender, M.L. Acc. Chem. Res., 1987 20 146. e) Breslow, R.; Czarnik, W. J. Amer. Chem. Soc., 1983, 105, 1390-1391; f) Tabushi, I.; Kuroda, Y.; Mochizuki, A. J. Amer. Chem. Soc., 1980, 102, 1152-1155; g) D'Souza, V.T.; Hanabusa, K.; O'Leary, T.; Gadwood, R.C.; Bender, M.L. Biochem. Biophys. Res. Comm., 1985, 129, 727. n) Ueno, A.; Suzuki, I.; Osa, T. J. Chem. Soc. Perkin Trans II, 1986, 1061. i) Fornasier, R.; Reniero, F.; Scrimin, P.; Tonellato, U. J. Chem. Soc., Perkin trans II 1987, 1121.
4. a) Ohkubo, K.; Miyake, S. J. Chem. Soc., Perkin trans II, 1987, 995. b) Murakami, Y.; Nakano, A.; Ikeda, H.; Imori, T.; Akiyoshi, K. Bull. Chem. Soc. Jpn. 1985, 58, 172.
5. a) Hosseini, M.W.; Lehn, J.M. J. Amer. Chem. Soc. 1982, 104, 3525-3527. b) Dietrich, B.; Fyles, D.L.; Fyles, T.M.; Lehn, J.M. Helv. Chim. Acta. 1979, 62 2763.
6. a) Cram, D.J. Science, 1974, 183 803. b) Lehn, J.M. Acc. Chem. Res. 1978, 11 49.
7. Odashima, K.; Koga, K. "Cyclophanes", Keehne, P.M.; Rosenfeld, S.M. Eds., Academic press, 1984, Vol. II.
8. a) Kim, M.; Gokel, G.W. J. Chem. Soc. Chem. Commun., 1987, 1686-1688. b) Feibush, B.; Saha, M.; Onan, K.; Karger, B.; Glese, R. J. Amer. Chem. Soc., 1987, 109, 7531-7533. c) Kelly, T.R.; Zhao, C.; Bridger, G.J. J. Amer. Chem. Soc., 1989, 111, 3744-3745.
9. (a) Youngquist, R.S.; Dervan, P.B. J. Amer. Chem. Soc. 1987, 109 7564-7566. (b) Rebek, J.; Askew, B.; Ballester, P.; Buhr, C.; Jones, S.; Nemeth, D.; Williams, K. *ibid.*, 1987, 109 5033. (c) Rebek, J.; Askew, B.; Ballester, P.; Buhr, C.; Costero, A.; Jones, S.; Williams, K. *ibid.*, 1987 109 6866. (d) Constant, J.F.; Fahy, J.; Lhomme, J. Tetrahedron Lett., 1987 28 1777. (e) Kelly, T.R.; Maquire, M.P. J. Amer. Chem. Soc. 1987, 109 6551-6553. (f) Seyama, F.; Akahori, K.; Sakata, Y.; Misumi, S.; Aida, M.; Nagata, C. J. Amer. Chem. Soc., 1988 110 2192-2201.
10. Schroeder, A.C.; Hughes, R.G.; Bloch, A. J. Amer. Chem. Soc., 1981, 103, 1078-1180.
11. Kelley, J.L.; Krochmal, M.P.; Schaeffer, H.J. J. Med. Chem., 1981, 24, 1528-1531.
12. Beauchamp, L.M.; Dolmatch, B.L.; Scheffer, H.J.; J. Med. Chem., 1985, 28 982-987.
13. Tso, P.O.P. in Basic Principles in Nucleic Acid Chemistry" Ed. Tso, P.O.P. Academic press (1974).